

REVIEW

Resveratrol and Cancer

Natalia G. Vallianou, MD, MSc, PhD,¹ Angelos Evangelopoulos, MSc, PhD,²
Eleni Geladari, MD,¹ Christos Kazazis, MD³

¹Department of Internal Medicine,
Evangelismos General Hospital, Athens,
Greece

²Roche Diagnostics Hellas, Athens,
Greece

³Samos, Greece

KEY WORDS: *resveratrol; apoptosis;
autophagy; anti-cancerous properties*

ABBREVIATIONS

AMPK = adenosine monophosphate-
activated protein kinase
COX = cyclooxygenase
CREB = cyclic AMP response element-
binding (protein)
ERK = extracellularly-regulated kinase(s)
MAPK = mitogen-activated protein
kinases
mTOR = mammalian target of rapamycin
nF-κB = nuclear factor kappa-light-chain-
enhancer of activated B cells
SIR = silent information regulator
STAT3 = signal transducer and activator
of transcription 3 (factor)
VEGF = vascular endothelial growth
factor

Address for correspondence:

Natalia G. Vallianou, MD,
5 Pyramidon Street, Marathonas,
Athens 190 05, Greece;
Tel.: +30 2294092359;
E-mail: natalia.vallianou@hotmail.com

Manuscript received February 5, 2015;

Revised manuscript received May 2,

2015; Accepted May 29, 2015

ABSTRACT

Resveratrol is a stilbene substance that belongs to the superfamily of phytoalexins, which are compounds synthesized by plants when stress occurs, such as during plant infection. It is abundant in red wine, red grapes, blueberries, peanuts and pistachios. Resveratrol induces p53-dependent apoptosis. A novel resveratrol analogue, HS-1793, has recently been demonstrated to inhibit vascular endothelial growth factor in human prostate cancer cells. Pterostilbene, an analog of resveratrol, has been demonstrated to exert both autophagy and apoptosis in human bladder and breast cancer cell lines. It has also been found to cause accumulation of autophagic vacuoles, as well as promote cell death via a mechanism involving lysosomal membrane permeabilization in human melanoma, colon, lung and breast cancer cell lines. Identification of a receptor site for resveratrol in cancer cells supports the potential of this compound as a therapeutic agent. The receptor could also serve as a vehicle for studies of future resveratrol analogues. Resveratrol has also been documented to overcome chemoresistance by inhibiting NF-κB and STAT3 pathway. Resveratrol has shown much promise in preclinical trials and because of its good safety profile it may be an ideal chemo-preventive and chemotherapeutic agent.

INTRODUCTION

Resveratrol (3,5,4'-trihydroxy-*trans*-stilbene) (Fig. 1) is a stilbene substance belonging to the superfamily of phytoalexins, which are compounds synthesized by plants in response to injury or stress, such as when the plant is infected by bacteria or fungi. It is abundant in red wine, in red grapes, blueberries, peanuts, and pistachios (Table 1).¹⁻³

Resveratrol exerts beneficial effects in humans and may be helpful in preventing and treating metabolic diseases, such as obesity, cardiovascular disease and diabetes mellitus.⁴⁻⁶ Resveratrol also possesses anti-oxidant and anti-cancerous properties, which will be discussed in this review.^{7,8} The anti-cancerous mechanisms of action of resveratrol are not well-understood, but it has been suggested from several studies that they are the result of resveratrol's action in inducing apoptosis.⁹⁻¹⁵

RESVERATROL AND p53

Resveratrol induces p53-dependent apoptosis.⁹⁻¹⁶ The p53 gene is a suppressive

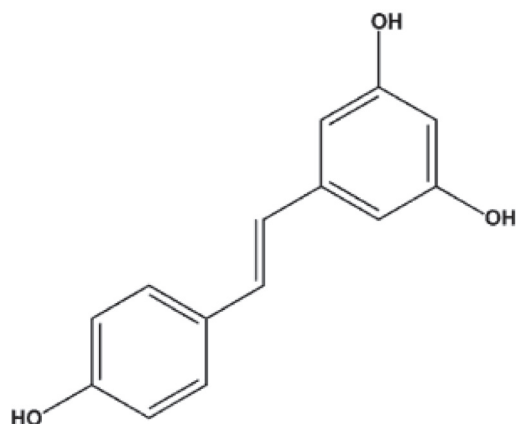


FIGURE 1. The chemical structure of resveratrol (3,5,4'-trihydroxy-trans-stilbene).

TABLE 1. Foods rich in resveratrol.

Red grapes
Red wine
Blueberries
Peanuts
Pistachios
Itadori tea
Sherries
Cranberry juice
Grape juice
Hops

oncogene, which exists at low concentrations in normal cells, whilst if there is DNA damage, levels of p53 activity rise, because of a post-translational mechanism that stabilizes the p53 encoded protein.¹⁷⁻¹⁹ p53 is involved in apoptosis and DNA repair.^{17,18} Phosphorylation of p53 occurs at some serine and threonine residues.^{19,20} Activated p53 binds to DNA by a mechanism, which depends on phosphorylation and acetylation of this protein. Phosphorylation of p53 at N-terminal site may promote stabilization of p53, and may be a factor that facilitates or is required for the acetylation of p53 at a C-terminal site. Acetylation has been reported to increase sequence-specific DNA binding of p53 *in vitro* and has been found to be essential for recruitment of the protein to coactivators—such as cyclic AMP response element-binding (CREB) protein/p300.²¹ These co-activators contain intrinsic histone acetyl-transferase properties. Acetylation of the p53 protein appears to be a related with p300/CREB-binding protein-associated factor (PCAF) (a histone acetyltransferase) and

several other p53 co-activators.²² It has also been suggested that resveratrol induces acetylation of p53. Phosphorylation of specific p53 residues is considered necessary for the activation of certain promoter regions.²³

Resveratrol-induced apoptosis factor (PCAF), another p53 co-activator, and CREB-binding protein/p300 are also co-activators for various transcription factors besides p53. Resveratrol is capable of phosphorylating p53 at N-terminal and C-terminal serine residues in many human cancer cell lines and also of promoting acetylation of p53. Activation of extracellularly-regulated kinases (ERK)-1 and -2 has been implicated in resveratrol-induced serine phosphorylation of p53 in cancer cells.⁹⁻¹⁵ It has been documented that resveratrol induces serine residue phosphorylation of a mutant p53 in prostate cancer cells, a phenomenon which induces apoptosis.^{9,10}

RESVERATROL AND CYCLOOXYGENASE (COX) - 2

Cyclooxygenase (COX), the enzyme involved in prostaglandin synthesis, is secreted by increased production of various inflammatory mediators. Two COX iso-enzymes have been described until now,^{24,25} COX-1 is a constitutively expressed form of the enzyme and is present everywhere, whereas COX-2 is inducible and is present in inflammatory lesions, and is known to be constitutively expressed in tumors. Constitutive expression of COX-2 in cells and animal models is associated with tumor cell growth and metastasis, enhanced cellular adhesion and inhibition of apoptosis.²⁶

COX-2 has been found in the endoplasmic reticulum, Golgi complex, and nuclear envelope.^{27,28} COX-1 and COX-2 are localized in the nuclear envelope and endoplasmic reticulum of prostaglandin-2 releasing cells. Recent data have documented that prostaglandin-2 biosynthesis, COX iso-enzymes, and prostaglandin-2 are located in the perinuclear region. Resveratrol induces nuclear accumulation of COX-2 in various cancer cells, such as human breast tumor cells, glioma, head and neck squamous cells, ovarian and prostate tumor cells.²⁸ These results suggest that inducible COX-2 may play a major role in p53-dependent apoptosis in tumor cells. Other researchers have also documented that COX-2 can be pro-apoptotic, while Hinz *et al* have demonstrated that COX-2 inhibitors could be deleterious for certain tumors because of the pro-apoptotic action of this protein.²⁹⁻³¹ Nevertheless, pharmacologic inhibition of COX-2 has resulted in conflicting results.^{32,33} Other data have suggested that over-expression of COX-2 could induce an anti-proliferative effect, which is attributed to p53 and p21 expression.^{34,35} Other studies support the notion that constitutive COX-2 expression induces tumor growth and is anti-apoptotic, whereas inducible COX-2, induced, e.g. by resveratrol and localized largely to the cell nucleus, is pro-

apoptotic by means of phosphorylation of the serine residue of p53. Such a mechanism may be unique for the treatment of many cancers. Clinical anti-cancer regimens could possibly be designed to target the constitutively expressed and inducible pools of COX-2, particularly for the management of tumors in which pharmacologic COX-2 inhibition produces cell cycle arrest, rather than apoptosis, and when resveratrol or other agents capable of inducing COX-2 result in apoptosis.^{36,37}

RESVERATROL AND SIRTUINS

The silent information regulator (SIR) genes (sirtuins) comprise a highly conserved family of proteins, with one or more sirtuins present in virtually all species, from prokaryotic organisms to eukaryotic ones. In mammals, 7 sirtuin genes -SIRT1 to SIRT7- have been identified.³⁸ There is emerging evidence that sirtuins constitute a very perplexed biological response system, which has a major impact on many other molecular pathways, such as aging, apoptosis and inflammation in complex manners. Resveratrol has been the first compound discovered, able to mimic calorie restriction by stimulating sirtuins.^{39,40} Calorie restriction is a process that alters the concentrations of many genes implicated in a variety of biological processes, such as growth, metabolism, immune system, as well as oxidative stress and DNA damage repair.⁴¹ The molecular effects induced by calorie restriction overlap with two major pathways linked with lifespan modulation *in vitro*, insulin/insulin-like growth factor signaling and target of rapamycin signaling.⁴¹ Calorie restriction is suggested to induce gene expression patterns in multiple tissues. Indeed, treatment with resveratrol has reduced tumorigenesis in SIRT1 +/-;p53 +/- mice, and this protective effect has been attributed to SIRT1 activation.⁴²

RESVERATROL AND EXTRACELLULAR SIGNAL-REGULATED KINASES (ERK) 1/2

Identification of a receptor site for resveratrol in cancer cells, by the implication of extracellular signal-regulated kinases 1 and 2 (ERK1/2) of the resveratrol signal downstream into p53-dependent apoptosis, supports the potential of this compound as a therapeutic agent.⁴¹ The receptor could also serve as a vehicle for studies of future resveratrol analogues.

Regarding tumor cells, the role of integrins is more complex than simply the transduction of outside in signals originating from integrin-matrix protein interactions. It is noteworthy that dysregulation of the β -3 integrins has been involved in the pathogenesis of cancer. Tumor growth and angiogenesis, such as those associated with vascular endothelial growth factor pathway, are enhanced in β -3-null mice.⁴³ On the con-

trary, integrin β -3 overexpression may suppress tumor growth of a human glioma model in rats.⁴⁴ The above-mentioned paradigms suggest that promotion of integrin β -3 expression in tumor cells could act as a therapeutic goal combating the process of carcinogenesis.

ERK1/2 are **mitogen-activated protein kinases** (MAPK) iso-enzymes, which serve as inducible components of the normal cellular signal transduction process. ERK1/2 activation pathway may be triggered in the setting of growth factor stimulated cells or by inflammation. MAPK-kinase activates ERK1/2 straight-forward.⁴⁵ Resveratrol activates MAPK at low concentrations, but higher concentrations of resveratrol can inhibit this signal transducing kinase in tumor cells.⁴⁶ It has been demonstrated that resveratrol induces ERK1/2 activation in prostate, breast, glial, head and neck, and ovarian cancer cells.⁴⁷ The activation of ERK1/2 by resveratrol may be blocked by a MAPK kinase or mitogen/extracellular signal-regulated kinase (MEK) inhibitor, PD98059.⁴⁸

RESVERATROL AND ADENOSINE MONOPHOSPHATE-ACTIVATED PROTEIN KINASE (AMPK)/MAMMALIAN TARGET OF RAPAMYCIN (MTOR) PATHWAY

Adenosine mono phosphate-activated protein kinase (AMPK) is linked with the phosphatidylinositol-3 kinase/AKT/mTOR signaling pathway, a cellular signaling cascade, which is of vital importance for cell growth, in response to mitogenic stimuli.^{49,50} AMP-activated protein kinase (AMPK) activation inhibits phosphorylation and activation of the mTORC1 complex and is partly controlled by the upstream kinase AKT (protein kinase B), whose activation decreases the AMP:ATP ratio.⁵¹ Resveratrol has also been shown to modulate AMPK in breakpoint cluster region protein (BCR)- Abelson murine leukemia viral oncogene homolog 1 (ABL 1) (BCR/ABL) gene transformed cells and to exhibit antileukemic effects.⁵³⁻⁵⁵ Treatment of either imatinib mesylate-sensitive or imatinib mesylate-resistant chronic myelogenous leukemia cells with resveratrol has resulted in apoptosis.⁵⁶

RESVERATROL, NUCLEAR FACTOR KAPPA-LIGHT-CHAIN-ENHANCER OF ACTIVATED B CELLS (NF- κ B) AND SIGNAL TRANSDUCER AND ACTIVATOR OF TRANSCRIPTION 3 (STAT3) FACTOR

The transcription factor, nuclear factor (NF)- κ B, regulates many genes implicated in growth regulation and inflammation.^{57,58} *In vitro* and *in vivo* studies have documented that constitutive activation of NF- κ B results in inhibition of chemotherapy-induced apoptosis in a number of cancer cells.⁵⁹⁻⁶⁴ Signal transducer and activator of transcription 3 (STAT3)

factor is also a ubiquitously expressed –like NF- κ B- member of the STAT family of transcription factors, which is activated by tyrosine phosphorylation by means of upstream receptors such as epidermal growth factor, platelet-derived growth factor and cytokines, for example interleukin-6.⁶⁵ Recent studies have demonstrated that STAT3 may confer cancer resistance to chemotherapeutic agents.⁶⁶⁻⁶⁹

STAT3 is one of the major compounds implicated in carcinogenesis.^{70,71} The oncogenic significance of activated STAT3 molecules is due to their effects on apoptosis, cell proliferation, angiogenesis, and immune system evasion.^{72,73} Constitutively active STAT3 has been involved in the promotion of resistance to apoptosis, probably through the expression of B-cell lymphoma-extra large (Bcl-xL) and cyclin D1 proteins.^{74,75} Its role in carcinogenesis is mediated through the induction of genes that suppress apoptosis and mediate proliferation and angiogenesis. Constitutive activation of STAT3 has been implicated in a variety of cancers, including breast, brain, colon, gastric, esophageal, ovarian, nasopharyngeal, pancreatic, prostate cancer, head and neck squamous cell carcinoma, multiple myeloma, lymphomas and leukemia.⁷⁶⁻⁷⁸ Nevertheless, it is not completely understood why STAT3 is constitutively active in cancer cells.

Resveratrol exerts its sensitization effect by modulating one or more mechanisms of chemo-resistance. Recent data have shown that resveratrol may overcome chemo-resistance in cancer cells by modulating apoptotic pathways, down-regulating drug transporters and down-modulating proteins involved in tumor cell proliferation. In addition, resveratrol has also been documented to overcome chemo-resistance by inhibiting NF- κ B and STAT3 pathway.^{79,80} Resveratrol has been suggested to enhance the apoptotic and anti-proliferative potential of bortezomib and thalidomide in multiple myeloma cells. Such an enhancement has been related to the inhibition of NF- κ B and STAT-3 activation pathways. Resveratrol administration has also been associated with accumulation of sub-G(1) population, increase in Bax release, and activation of caspase-3. This has been further related with down-regulation of various proliferative and anti-apoptotic gene products, including cyclin D1, cellular inhibitor of apoptosis 2 (cIAP-2), X-linked inhibitor of apoptosis protein (XIAP), survivin, B-cell lymphoma 2 (Bcl-2), Bcl-xL, Bfl-1/A1, and tumor necrosis factor receptor-associated factor 2 (TRAF2).⁸¹ Investigation of the mechanism has revealed that resveratrol inhibited NF- κ B activation through inhibition of I κ B α phosphorylation and (I κ B kinase) IKK activation. These observations have been further supported by an inhibition of NF- κ B and STAT-3 in patients with multiple myeloma.⁸²

RESVERATROL AND AUTOPHAGY

Autophagy is an evolutionarily conserved intracellular

process, characterized by lysosomal degradation of proteins, which is essential for survival of eukaryotic cells under metabolic stress. It has also been suggested to act as a form of programmed cell death.⁸³⁻⁸⁶ Pterostilbene, an analog of resveratrol, has been demonstrated to exert both autophagy and apoptosis in human bladder and breast cancer cell lines.^{87, 88} It has also been found to cause accumulation of autophagic vacuoles as well as promote cell death *via* a mechanism involving lysosomal membrane permeabilization in human melanoma, colon, lung and breast cancer cell lines.^{89, 90} Pterostilbene has been documented to produce autophagy in human leukemia cells.⁹¹

RESVERATROL, VASCULAR ENDOTHELIAL GROWTH FACTOR (VEGF) AND PI3K/Akt

A novel resveratrol analogue, HS-1793, has recently been demonstrated to inhibit vascular endothelial growth factor (VEGF) in human prostate cancer cells. HS-1793 has been suggested to inhibit phosphorylation of PI3K and Akt in human prostate cancer cells.⁹² Resveratrol itself has been suggested to inhibit the PI3K and Akt pathway in acute lymphoblastic leukemia cells.⁹³ Also, it has substantially induced HIF-1 α protein degradation by means of the proteasome pathway. Moreover, HS-1793 has shown more potent effects than resveratrol on the cytotoxic effects on PC-3 cells.⁹⁴ Resveratrol has been found to possess anti-angiogenic properties, through the inhibition of VEGF, useful for the prevention of breast cancer, too.⁹⁵ Also, this resveratrol analogue, HS-1793, has been shown to induce cell cycle arrest and apoptotic cell death, in human breast cancer cells. In particular, it has been found to induce G2/M cell arrest in human breast cancer cells.⁹⁶

RESVERATROL AND ATF3 TRANSCRIPTION FACTOR

Recently, ATF3 has been identified as a novel target of resveratrol in colorectal cancer cells.⁹⁷ ATF3, a member of the ATF/CREB family of transcription factors, is characterized as an adaptive response gene.⁹⁸ Latest data suggest that ATF3 may function as a tumor suppressor gene in colorectal tumorigenesis. First, ATF3 expression is substantially reduced in cancer tissues, compared to normal tissue.⁹⁹ Second, ATF3 over-expression is reported to produce inhibition of proliferation, promotion of apoptosis, inhibition of invasion and decrease of tumor formation *in vivo*.¹⁰⁰⁻¹⁰³ Finally, ATF3 is demonstrated to enhance induction of apoptosis by substances known to possess anti-cancerous properties.¹⁰⁴⁻¹⁰⁶ Thus, it is suggested that ATF3 plays an anti-carcinogenic role in colorectal cancer (Fig. 2).¹⁰⁷

BIOAVAILABILITY OF RESVERATROL

Resveratrol's bioavailability is compromised by its physicochemical properties, such as low stability, increased oxidation on heat and light exposure, low water solubility as well as its high hepatic uptake. Data obtained from human pharmacokinetic studies have shown a low amount of intact resveratrol in the systemic circulation, which does not justify its therapeutic activities, raising doubts about resveratrol's potential *in vivo*.¹⁰⁸ Recently, a soluble form of trans-resveratrol has been developed, which has been demonstrated to be better absorbed and to have efficient serum levels compared to the dry powder. A single dose of 40 mg of this soluble galenic form resulted in blood levels of 0.1-6 μ M for several hours and without any observed intolerance or toxicity.¹⁰⁹

CONCLUSION

Resveratrol has shown much promise in preclinical trials and because of its good safety profile it may be an ideal chemopreventive and chemotherapeutic agent. However, the rapid metabolism of resveratrol has been a continuing challenge. Researchers now are focusing on approaches to overcome this problem, which appears to be a major obstacle in the clinical use of resveratrol. However, resveratrol seems to have a long way ahead until it could find its place as an effective chemotherapeutic agent.

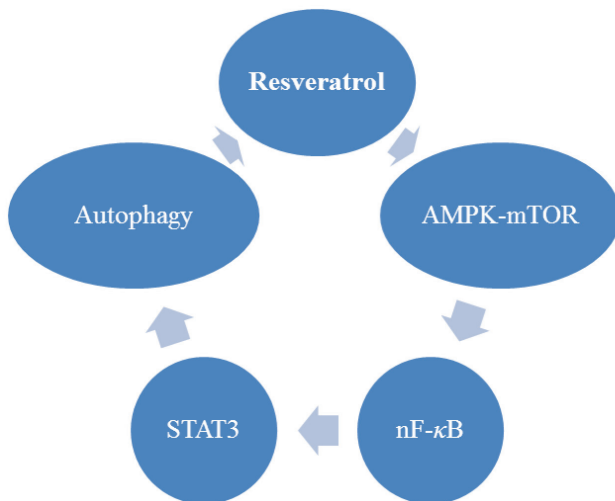


FIGURE 2. The main different molecular pathways through which resveratrol exerts its anti-cancerous effects. AMPK = 5' adenosine monophosphate-activated protein kinase; mTOR = mammalian target of rapamycin; nF-kB = nuclear factor kappa-light-chain-enhancer of activated B cells; STAT3 = signal transducer and activator of transcription 3 (factor).

REFERENCES

- Baur JA, Sinclair DA. Therapeutic potential of resveratrol: The *in vivo* evidence. *Nat Rev Drug Dis* 2006; 5:493-506.
- Burns J, Yokota T, Ashihara H, Lean MJ, Crozier A. Plant foods and herbal sources of resveratrol. *J Agric Food Chem* 2002; 50:3337-3340.
- Langecake P, Pryce RJ. The production of resveratrol by *Vitis vinifera* and other members of the Vitaceae as a response to infection or injury. *Physiol Plant Pathol* 1976; 9:77-86.
- Frojdo S, Durand C, Pirola L. Metabolic effects of resveratrol in mammals—a link between improved insulin action and aging. *Curr Aging Sci* 2008; 1:145-151.
- Szkudelska K, Szkudelski T. Resveratrol, obesity and diabetes. *Eur J Pharmacol* 2010; 635:1-8.
- Su HC, Hung LM, Cheng JK. Resveratrol, a red wine antioxidant, possesses an insulin-like effect in streptozotocin-induced diabetic rats. *Am J Physiol Endocrinol Metab* 2006; 290:1339-1346.
- Athar M, Back JH, Kopelovich L, Bickers DR, Kim AL. Multiple molecular targets of resveratrol: Anti-carcinogenic mechanisms. *Arch Biochem Biophys* 2009; 486:95-102.
- Jang M, Cai L, Udeani GO, et al. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science* 1997; 275:218-220.
- Lin HY, Shih A, Davis FB, et al. Resveratrol-induced serine phosphorylation of p53 causes apoptosis in a mutant p53 prostate cancer cell line. *J Urol* 2002; 168:748-755.
- Shih A, Zhang SL, Cao JH, et al. Inhibitory effect of EGF on resveratrol-induced apoptosis in prostate cancer cells is mediated by protein kinase C- α . *Mol Cancer Ther* 2004; 11:1355-1364.
- Zhang SL, Davis FB, Cao JH, et al. Y. Oestrogen inhibits resveratrol-induced posttranslational modification of p53 and apoptosis in breast cancer cells. *Br J Cancer* 2004; 91:178-185.
- Tang HY, Shih A, Cao JH, et al. Inducible COX-2 facilitates p53-dependent apoptosis in human breast cancer cells. *Mol Cancer Ther* 2006; 5:2034-2040.
- Lin HY, Tang HY, Keating T, et al. Resveratrol is pro-apoptotic and thyroid hormone is anti-apoptotic in glioma cells: both actions are integrin- and ERK-mediated. *Carcinogenesis* 2008; 29:62-69.
- Lin HY, Sun MZ, Tang HY, et al. Resveratrol causes COX-2- and p53-dependent apoptosis in head and neck squamous cell cancer cells. *J Cell Biochem* 2008; 104:2131-2142.
- Lin C, Crawford D, Lin S, et al. Inducible COX-2-dependent apoptosis in human ovarian cancer cells. *Carcinogenesis* 2011; 32:19-26.
- Mahyar-Roemer M, Roemer K. p21Waf1/Cip1 can protect human colon carcinoma cells against p53-dependent and p53-independent apoptosis induced by natural chemopreventive and therapeutic agents. *Oncogene* 2001; 20:3387-3398.
- Lavin MF, Gueven N. The complexity of p53 stabilization and activation. *Cell Death Differ* 2006; 13:941-950.
- Agarwal ML, Taylor WR, Chernov MV, Chernova OB, Stark GR. The p53 network. *J Biol Chem* 1998; 273:1-4.

19. Colman MS, Afshari CA, Barrett JC. Regulation of p53 stability and activity in response to genotoxic stress. *Mutat Res* 2000; 462:179-188.
20. Kapoor M, Hamm R, Yan W, Taya Y, Lozano G. Cooperative phosphorylation at multiple sites is required to activate p53 in response to UV radiation. *Oncogene* 2000; 19:358-364.
21. Barlev NA, Liu L, Chehab NH, et al. Acetylation of p53 activates transcription through recruitment of coactivators/histone acetyltransferases. *Mol Cell* 2001; 8:1243-1254.
22. Dornan D, Shimizu H, Perkins ND, Hupp TR. DNA-dependent acetylation of p53 by the transcription coactivator p300. *J Biol Chem* 2003; 278:13431-13441.
23. Appella E, Anderson CW. Post-translational modifications and activation of p53 by genotoxic stresses. *Eur J Biochem* 2001; 268:2764-2772.
24. Morita I, Schindler MK Regier, et al. Different intracellular locations for prostaglandin endoperoxide H synthase-1 and -2. *J Biol Chem* 1995; 270:10902-10908.
25. Smith WL, Garavito RM, DeWitt DL. Prostaglandin endoperoxide-synthases (cyclooxygenases)-1 and -2. *J Biol Chem* 1996; 271:33157-33160.
26. Nakata R, Takahashi S, Inoue H. Recent Advances in the Study on Resveratrol. *Biol Pharm Bull* 2012; 35:273-279.
27. Murakami M, Nakashima D, Masuda S, et al. Cellular prostaglandin E2 production by membrane-bound prostaglandin E synthase-2 via both cyclooxygenases-1 and -2. *J Biol Chem* 2003; 278:37937-37947.
28. Parfenova H, Parfenov BV, Levine V, et al. Dynamics of nuclear localization sites for COX-2 in vascular endothelial cells. *Am J Physiol Cell Physiol* 2001; 281:C166-C178.
29. Ueno NM, Murakami T, Tanioka Fujimori K, et al. Coupling between cyclooxygenase, terminal prostanoid synthase, and phospholipase A2. *J Biol Chem* 2001; 276:34918-34927.
30. Zahner GG, Wolf R, Reinking R, et al. Cyclooxygenase-2 overexpression inhibits platelet-derived growth factor-induced mesangial cell proliferation through induction of the tumor suppressor gene p53 and the cyclin-dependent kinase inhibitors p21waf-1/cip-1 and p27kip-1. *J Biol Chem* 2002; 277:9763-9771.
31. Yuan B, Ohyama K, Bessho T, Toyoda H. Contribution of inducible nitric oxide synthase and cyclooxygenase-2 to apoptosis induction in smooth chorion trophoblast cells of human fetal membrane tissues. *Biochem Biophys Res Commun* 2006; 341:822-827.
32. Hinz B, Ramer R, Eichele K, Weinzierl U, Brune K. Up-regulation of cox-2 expression is involved in R(+)-methanandamide-induced apoptosis of human neuroglioma cells. *Mol Pharmacol* 2004; 66:1643-1651.
33. Han JA, Kim JI, Ongusaha PP, et al. p53 mediated induction of Cox-2 counteracts p53- or genotoxic stress-induced apoptosis. *EMBO J* 2002; 21:5635-5644.
34. Moalic-Juge S, Liagre B, Duval R, et al. The antiapoptotic property of NS-398 at high dose can be mediated in part through NF-kappaB activation, hsp70 induction and a decrease in caspase-3 activity in human osteosarcoma cells. *Int J Oncol* 2002; 20:1255-1262.
35. Swamy MV, Herzog CR, Rao CV. Inhibition of COX-2 in colon cancer cell lines by celecoxib increases the nuclear localization of active p53. *Cancer Res* 2003; 63:5239-5242.
36. Williams JL, Nath N, Chen J, et al. Growth inhibition of human colon cancer cells by nitric oxide Resveratrol-induced apoptosis (NO)-donating aspirin is associated with cyclooxygenase-2 induction and beta-catenin/T-cell factor signaling, nuclear factor-kappaB, and NO synthase 2 inhibition: implications for chemoprevention. *Cancer Res* 2002; 63:7613-7618.
37. King JG Jr, Khalili K. Inhibition of human brain tumor cell growth by the anti-inflammatory drug, flurbiprofen. *Oncogene* 2001; 20:6864-6870.
38. Kelly GS. A review of the sirtuin system, its clinical implications, and the potential role of dietary activators like resveratrol. *Altern Med Rev* 2010; 15:313-328.
39. Huang J, Plass C, Gerhauser C. Cancer chemoprevention by targeting the epigenome. *Curr Drug Targets* 2011; 12:1925-1956.
40. Howells LM, Berry DP, Elliott PJ, Jacobson EW. Phase I randomized, double-blind pilot study of micronized resveratrol (SRT501) in patients with hepatic metastases--safety, pharmacokinetics, and pharmacodynamics. *Cancer Prev Res (Phila)* 2011; 4:1419-1425.
41. Vang O, Ahmad N, Baile CA, Baur JA. What is new for an old molecule? Systematic review and recommendations on the use of resveratrol. *PLoS One* 2011; 6:e19881.
42. Zhao G, Guo S, Somel M, Khaitovich P. Evolution of human longevity uncoupled from calorie restriction mechanisms. *PLoS One* 2014; 9:e84117.
43. Villalba JM, Alcain FJ. Sirtuin activators and inhibitors. *Biofactors* 2012; 38:349-359.
44. Kanamori M, Vanden Berg SR, Bergers G, Berger MS, Pieper RO. Integrin beta3 overexpression suppresses tumor growth in a human model of gliomagenesis: implications for the role of beta3 overexpression in glioblastoma multiforme. *Cancer Res* 2004; 64:2751-2758.
45. Lin HY, Tang HY, Davis FB, Davis PJ. Resveratrol and apoptosis. *Ann New York Acad Sci* 2011; 1215:79-88.
46. Bergh JJ, Lin HY, Lansing L, et al. L-Thyroxine induces mitogen-activated protein kinase activation via binding to integrin alpha(V)beta3. *Endocrinology* 2005; 146:2864-2871.
47. Lassus P, Roux P, Zugasti O, et al. Extinction of rac1 and Cdc42Hs signalling defines a novel p53-dependent apoptotic pathway. *Oncogene* 2000; 19:2377-2385.
48. Dong Z. Molecular mechanism of the chemopreventive effect of resveratrol. *Mutat Res* 2003; 523-524:145-150.
49. Hardie DG. AMPK and Raptor: matching cell growth to energy supply. *Mol Cell* 2008; 30: 263-265.
50. Hahn-Windgassen A, Nogueira V, Chen CC, et al. Akt activates the mammalian target of rapamycin by regulating cellular ATP level and AMPK activity. *J Biol Chem* 2007; 280:32081-32089.
51. Wu Y, Liu F. Targeting mTOR: evaluating the therapeutic potential of resveratrol for cancer treatment. *Anticancer Agents Med Chem* 2013; 13:1032-1038.

52. Gupta SC, Sundaram C, Reuter S, Aggarwal BB. Inhibiting NF-kappaB activation by small molecules as a therapeutic strategy. *Biochim Biophys Acta* 2010; 10-12:775-787.
53. Sen R, Baltimore D. Multiple nuclear factors interact with the immunoglobulin enhancer sequences. *Cell* 1986; 46:705-716.
54. Puissant A, Grosso S, Jacquelin A, et al. Imatinib mesylate-resistant human chronic myelogenous leukemia cell lines exhibit high sensitivity to the phytoalexin resveratrol. *Faseb J* 2008; 22:1894-1904.
55. Puissant A, Robert G, Fenouille N, et al. Resveratrol promotes autophagic cell death in chronic myelogenous leukemia cells via JNK-mediated p62/SQSTM1 expression and AMPK activation. *Cancer Res* 2010; 70:1042-1052.
56. Vacana E, Plataniotis LC. AMPK in BCR-ABL expressing leukemias. Regulatory effects and therapeutic implications. *Oncotarget* 2012; 2:1322-1328.
57. Bharti AC, Aggarwal BB. Chemopreventive agents induce suppression of nuclear factor-kappa B leading to chemosensitization. *Ann NY Acad Sci* 2002; 973:392-395.
58. Bharti AC, Aggarwal BB. Nuclear factor-kappa B and cancer: its role in prevention and therapy. *Biochem Pharmacol* 2002; 64:883-888.
59. Wang CY, Mayo MW, Baldwin AS Jr. TNF- and cancer therapy-induced apoptosis: potentiation by inhibition of NF-kappa B. *Science* 1996; 274:784-787.
60. Ghiringhelli F, Rebe C, Hichami A, Delmas D. Immunomodulation and anti-inflammatory roles of polyphenols as anticancer agents. *Anticancer Agents Med Chem* 2012; 12:852-873.
61. Spatafora C, Tringali C. Natural-derived polyphenols as potential anticancer agents. *Anticancer Agents Med Chem* 2012; 12:902-918.
62. Fulda S. Regulation of cell death and survival by resveratrol: implications for cancer therapy. *Anticancer Agents Med Chem* 2012; 12:874-879.
63. Levy DE, Darnell JE Jr. Stats: transcriptional control and biological impact. *Nat Rev Mol Cell Biol* 2002; 3:651-662.
64. Bharti Weber D, Alexanian R, Raj-Vadhan S, et al. Nuclear factor-kappaB and STAT3 are constitutively active in CD138+ cells derived from multiple myeloma patients, and suppression of these transcription factors leads to apoptosis. *Blood* 2004; 103:3175-3184.
65. Real PJ, Sierra A, De Juan A, et al. Resistance to chemotherapy via Stat3-dependent overexpression of Bcl-2 in metastatic breast cancer cells. *Oncogene* 2002; 21:7611-7618.
66. Bhardwaj A, Sethi G, Vadhan-Raj S, et al. Resveratrol inhibits proliferation, induces apoptosis, and overcomes chemoresistance through down-regulation of STAT3 and nuclear factor kappaB-regulated antiapoptotic and cell survival gene products in human multiple myeloma cells. *Blood* 2007; 109:2293-2302.
67. Ahn KS, Sethi G, Sung B, et al. Guggulsterone, a farnesoid X receptor antagonist, inhibits constitutive and inducible STAT3 activation through induction of a protein tyrosine phosphatase SHP-1. *Cancer Res* 2008; 68:4406-4415.
68. Gamero AM, Young HA, Wiltrout RH. Inactivation of Stat3 in tumor cells: releasing a brake on immune responses against cancer? *Cancer Cell* 2004; 5:111-112.
69. Bowman T, Garcia R, Turkson J, Jove R. STATs in oncogenesis. *Oncogene* 2000; 19:2474-2488.
70. Kortylewski M, Kujawski M, Wang T, et al. Inhibiting Stat3 signaling in the hematopoietic system elicits multicomponent antitumor immunity. *Nat Med* 2005; 11:1314-1321.
71. Catlett-Falcone R, Landowski TH, Oshiro MM, et al. Constitutive activation of Stat3 signaling confers resistance to apoptosis in human U266 myeloma cells. *Immunity* 1999; 10:105-115.
72. Zushi S, Shinomura Y, Kiyohara T, et al. STAT3 mediates the survival signal in oncogenic ras-transfected intestinal epithelial cells. *Int J Cancer* 1998; 78:326-330.
73. Huang M, Page C, Reynolds RK, Lin J. Constitutive activation of stat 3 oncogene product in human ovarian carcinoma cells. *Gynecol Oncol* 2000; 79:67-73.
74. Roder S, Steimle C, Meinhardt G, Pahl HL. STAT3 is constitutively active in some patients with polycythemia rubra vera. *Exp Hematol* 2001; 29:694-702.
75. Mora LB, Buettner R, Seigne J, et al. Constitutive activation of Stat3 in human prostate tumors and cell lines: direct inhibition of Stat3 signaling induces apoptosis of prostate cancer cells. *Cancer Res* 2002; 62:6659-6666.
76. Greten FR, Weber CK, Greten TF, et al. Stat3 and NF-kappaB activation prevents apoptosis in pancreatic carcinogenesis. *Gastroenterology* 2002; 123:2052-2063.
77. Kirito K, Nagashima T, Ozawa K, Komatsu M. Constitutive activation of Stat1 and Stat3 in primary erythroleukemia cells. *Int J Hematol* 2002; 75:51-54.
78. Schaefer LK, Ren Z, Fuller GN, Schaefer TS. Constitutive activation of Stat3alpha in brain tumors: localization to tumor endothelial cells and activation by the endothelial tyrosine kinase receptor (VEGFR-2). *Oncogene* 2002; 21:2058-2065.
79. Hsiao JR, Jin YN, Tsai ST, Shiao AL, Wu CL, Su WC. Constitutive activation of STAT3 and STAT5 is present in the majority of nasopharyngeal carcinoma and correlates with better prognosis. *Br J Cancer* 2003; 89:344-349.
80. Song L, Turkson J, Karras JG, Jove R, Haura EB. Activation of Stat3 by receptor tyrosine kinases and cytokines regulates survival in human non-small cell carcinoma cells. *Oncogene* 2003; 22:4150-4165.
81. To KF, Chan MW, Leung WK, et al. Constitutional activation of IL-6-mediated JAK/STAT pathway through hypermethylation of SOCS-1 in human gastric cancer cell line. *Br J Cancer* 2004; 91:1335-1341.
82. Lin Q, Lai R, Chirieac LR, et al. Constitutive activation of JAK3/STAT3 in colon carcinoma tumors and cell lines: inhibition of JAK3/STAT3 signaling induces apoptosis and cell cycle arrest of colon carcinoma cells. *Am J Pathol* 2005; 167:969-980.
83. Gupta SC, Kannapann R, Reuter S, Kim JH, Aggarwal BB. Chemosensitization of tumors by resveratrol. *Ann N Y Acad Sci* 2011; 150-160:2011.
84. Kroemer G, Levine B. Autophagic cell death: the story of a misnomer. *Nat Rev Mol Cell Biol* 2008; 9:1004-1010.
85. Eskelinen EL. Maturation of autophagic vacuoles in Mammalian cells. *Autophagy* 2005; 1:1-10.

86. Chen RJ, Ho CT, Wang YJ. Pterostilbene induces autophagy and apoptosis in sensitive and chemoresistant human bladder cancer cells. *Mol Nutr Food Res* 2010; 54:1819-1832.
87. Wang Y, Ding L, Wang X. Pterostilbene simultaneously induces apoptosis, cell cycle arrest and cyto-protective autophagy in breast cancer cells. *Am J Transl Res* 2012; 4:44-51.
88. Mena S, Rodriguez ML, Ponsoda X, et al. Pterostilbene-induced tumor cytotoxicity: a lysosomal membrane permeabilization-dependent mechanism. *PLoS One* 2012; 7:e44524.
89. Wang Q, Mora-Jensen H, Weniger MA, Perez-Galan P, Wolford C. ERAD inhibitors integrate ER stress with an epigenetic mechanism to activate BH3-only protein NOXA in cancer cells. *Proc Natl Acad Sci* 2009; 106:2200-2205.
90. Kapetanovic IM, Muzzio M, Huang Z, Thompson TN, McCormick DL. Pharmacokinetics, oral bioavailability, and metabolic profile of resveratrol and its dimethylether analog, pterostilbene, in rats. *Cancer Chemother Pharmacol* 2011; 68:593-601.
91. Siedlecka-Kroplewska K, Jozwik A, Boguslawski W, et al. Pterostilbene induces accumulation of autophagic vacuoles followed by cell death in HL60 human leukemia cells. *J Physiol Pharmacol* 2013; 64:545-556.
92. Kim DH, Hossain MA, Kim MY, et al. A novel resveratrol analogue, HS-1793 inhibits hypoxia-induced HIF-1 α and VEGF expression and migration in human prostate cancer cells. *Int J Oncol* 2013; 43:1915-1924.
93. Li Y, Wicha MS, Schwartz SJ, Sun D. Implication of cancer stem cell theory for cancer chemoprevention by natural dietary compounds. *J Nutr Biochem* 2011; 22:799-806.
94. Liu MM, Huang Y, Yang J. Developing phytoestrogens for breast cancer prevention. *Anticancer Agents Med Chem* 2012; 12:1306-1313.
95. Kim JA, Kim DH, Hossain MA, et al. HS-1793, a resveratrol analogue, induces cell cycle arrest and apoptotic cell death in human breast cancer cells. *Int J Oncol* 2014; 44:473-480.
96. Whitlock NC, Bahn JH, Lee SH, Eling TE, Baek SJ. Resveratrol-induced apoptosis is mediated by early growth response-1, kröppel-like factor 4, and activating transcription factor 3. *Cancer Prev Res* 2011; 4:116-127.
97. Lu D, Chen J, Hai T. The regulation of ATF3 gene expression by mitogen-activated protein kinases. *Biochem J* 2007; 401:559-565.
98. Yan C, Boyd DD. ATF3 regulates the stability of p53: a link to cancer. *Cell Cycle* 2006; 5:926-929.
99. Fan F, Jin S, Amundson SA, Tong T, Fan W. ATF3 induction following DNA damage is regulated by distinct signaling pathways and over-expression of ATF3 protein suppresses cell growth. *Oncogene* 2002; 21:7488-7495.
100. Huang X, Li X, Guo B. KLF6 induces apoptosis in prostate cancer cells through up-regulation of ATF3. *J Biol Chem* 2008; 283:29795-29801.
101. Lu D, Wolfgang CD, Hai T. Activating transcription factor 3, a stress-inducible gene, suppresses Ras-stimulated tumorigenesis. *J Biol Chem* 2006; 281:10473-10481.
102. Yamaguchi K, Lee SH, Kim JS, Wimalasena J, Kitajima S. Activating Transcription factor 3 and early growth response 1 are the novel targets of LY294002 in a phosphatidylinositol 3-kinase-independent pathway. *Cancer Res* 2006; 66:2376-2384.
103. Turchi L, Aberdam E, Mazure N, Pouyssegur J, Deckert M. Hif-2 α mediates UV-induced apoptosis through a novel ATF3-dependent death pathway. *Cell Death Differ* 2008; 15:1472-1480.
104. Bottone FG, Moon Y, Kim JS, Alston-Mills B, Ishibashi M. The anti-invasive activity of cyclooxygenase inhibitors is regulated by the transcription factor ATF3 (activating transcription factor 3). *Mol Cancer Ther* 2005; 4:693-703.
105. Whitlock NC, Baek SJ. The anti-cancer effects of resveratrol-Modulation of transcription factors. *Nutr Cancer* 2012; 64:493-502.
106. Schlessinger K, Levy DE. Malignant transformation but not normal cell growth depends on signal transducer and activator of transcription 3. *Cancer Res* 2005; 65:5828-5834.
107. Masuda M, Suzui M, Yasumatu R, et al. Constitutive activation of signal transducers and activators of transcription 3 correlates with cyclin D1 overexpression and may provide a novel prognostic marker in head and neck squamous cell carcinoma. *Cancer Res* 2002; 62:3351-3355.
108. Santos AC, Veiga F, Ribeiro AJ. New delivery systems to improve the bioavailability of resveratrol. *Expert Opin Drug Deliv* 2011; 8:973-990.
109. Amiot MJ, Romier B, Dao TM, et al. Optimization of trans-Resveratrol bioavailability for human therapy. *Biochimie* 2013; 95:1233-1238.